Summary

Investigations carried out for this study were based on three main objectives. The primary objective of this study was to isolate and enumerate mercury-resistant bacteria and to compare their occurrence and distribution spatially and temporally along the Indian coasts. As coastal waters experience adverse effects of a variety of toxic chemicals and salt concentration that may play a role in resistance mechanism in mercury-resistant marine bacteria, investigations were carried out to see whether marine MRB have different ways to deal with toxic mercury and to examine their genetic make-up. The hypothesis put forth for this study that bacterial strains capable of Hg resistance can also tolerate, detoxify or biotransform a variety of other toxicants was examined. Experiments were carried out to understand the potential of a select set of MRB to tolerate and/or biotransform polychlorinated biphenyls (PCBs), tri-butyl tin (TBT), mercury, cadmium and lead to validate this hypothesis.

Enumeration of MRB was carried out from water and sediments over a period of four years (1999-2002) and comparison was made with previous records (1993-1997). Thirty randomly isolated environmental strains of these MRB were characterized biochemically. In addition, genomic DNA from a select set of 13 strains of MRB was investigated for presence of *mer* operon and six of its structural genes and also for occurrence of plasmids. Upon selecting many MRB strains that were able to grow in the media containing many of these toxicants individually. Further detailed analyses involved quantification of detoxification/removal of toxicants such as heavy metals (Cd, Pb), PCBs and TBT with a view to understand their detoxification potential and applicability of these marine MRB for bioremediation.

Results and inferences from these investigations can be summarized as follows.

Contamination caused by heavy metals like mercury affects both oceanic, continental shelf and coastal regimes. In the latter zones, besides a longer residence time, metal concentrations are

higher due to perpetual input and transport by river runoff and from nearby industrial and urban zones. The spatial and temporal variation of MRB along the Indian coasts over a period of nine (1993-2002) years described in this thesis give a clear view of the scenario of abundance of MRB and indirectly indicates the growing menace of heavy metal pollution in the coastal environments. **Recent increase in MRB** is important considering the present status of mercury consumption in India. Staggering abundances of MRB are useful not only to ascertain the recent changes the coastal environs of the country have gone through in terms of Hg increases, but also, as a reliable practical method, their enumeration will prove useful, akin to other indicators, the extent of Hg and other heavy metals pollution. This study supports the fact that heavy metalresistance does not arise by chance, rather there has to be a selection pressure from natural and/or anthropogenic inputs to bring this change. The fact that India holds currently the numero uno position in the list of mercury-consuming countries, is indicative enough in ascertaining as to why there was an unusual rise in MRB along the coast post 1999. In that, while MRB were rarely found from samples analyzed until 1997 using seawater nutrient agar with 20 times lesser concentrations of Hg, their occurrence from a minimum 13% to as high as 100% of samples analyzed from 1999 onwards. Corroborating quite well with this observation is the increase in mercury concentration in the environment as reported by Central Pollution Control Board and others. The fact that a major portion of the natural, culturable bacterial flora was mercuryresistant from the offshore regions of Bay of Bengal (BOB) does point to the global nature of the pollution problem caused by this most toxic heavy metal. It is also important to note that the presence of higher percentages of MRB in the offshore waters of the BOB might imply that in these low salinity areas, there is an adverse impact of heavy metals on the overall biological productivity. Presence of higher counts of MRB in the deeper waters from both of the coastal and oceanic locations of the BOB does suggest that a greater portion of bacterial community is resistant to this heavy metal. Upon collecting reasonably large amounts of environmental data on distribution of MRB along the Indian coasts, it is affirmative that the MRB have adapted to the coastal environments vulnerable to pollution threats and consequences. It was then hypothesized that the MRB must be possessing mechanism(s) to deal with not only Hg, but several other toxic pollutants including heavy metals and xenobiotics.

A total of 83 mercury-resistant bacteria from different coastal environments of India were isolated on selective SWNA medium amended with 10 ppm mercury. Thirty of these bacteria were submitted to an extensive biochemical characterization protocols by following MacFaddin (1980). Thirteen of these thirty isolates which tolerated mercury concentration ≥ 25 ppm, termed as bacteria highly resistant to mercury (BHRM), were subjected to 16S rDNA sequencing. Seven out of these13 isolates were found to be Alcaligenes faecalis. Four were Bacillus sp., one Pseudomonas and one Brevibacterium iodinium. Marked differences in identification using biochemical characterization and 16S rDNA sequence suggests that more reliable biochemical tests need to be established for identification of marine bacteria in general and MRB, in particular. The resistance of MRB was always quite remarkable and they were able to detoxify/degrade a variety of toxic chemicals. Their growth was invariably diauxic in the presence of toxic chemicals examined during this study. More importantly, there was a synergistic effect of combined toxicity i.e. requiring lower concentrations in combination to stop the MRB growth. These bacteria were highly resistant to an array of antibiotics conforming to the fact that most of the times heavy metal- resistance is associated with antibiotic resistance. Though not experimented in a great details, analysis of C:N ratio as a measure of understanding the mechanism of mercury resistance during this study is quite important to suggest that underlying resistant mechanism in MRB is not always universal; one or more pathways operate and that, possibly, not all regulatory and physiological mechanisms are required for dealing with Hg toxicity.

The results from the analyses of *mer* genes from the 11 bacteria were very interesting. Nine of them had *merA* (important in reducing Hg^{2+} to Hg^{0}). Four of these isolates possessed *merB*. Whereas, merD was present in three isolates. All of these 11 isolates were seen to volatilise mercury. MerR gene was found in four isolates. BHRM strains, CH13 (Bacillus sp.) and CM10 (Alcaligenes faecalis) though negative for merA, were able to volatilize Hg. Absence of transport proteins like merP, merC as evidenced from the absence of merP or merC genes, in the marine MRB strains examined during this study calls for investions on different mechanisms of mercury transport. It is also likely that mer gene arrangement in the marine strains has a different orientation as the universal mer oligomers failed to amplify total mer from all the 11 isolates examined through PCR during this study. It is hypothesized from the molecular analyses that, probably the role of Na⁺ in transport of Hg²⁺ into the cell and role of Cl⁻ in determining mercury bioavailability are of greater importance in the marine strains of MRB. Moreover, recognition of non-mer mediated mercury volatilization during this study is of great importance in understanding the detoxification mechanisms offered by several mercury-resistant marine bacteria. Absence of plasmids, as confirmed both by molecular analyses and by two indirect assays prompts to conclude that mercury resistance in marine bacteria is largely chromosomemediated. All these findings are interesting and prompted some research on their detoxification mechanisms. While what has been achieved by this study is highly promising, there is a need for more detailed investigations to exploit their potential in detoxification of heavy metals including Hg.

These bacteria were capable of growth in medium amended with various heavy metals like Hg, Cd and Pb. All the eleven mercury-resistant bacteria tested were efficient in volatilizing mercury from a buffered solution containing 10 ppm mercury. Isolate CH07 identified as Pseudomonas aeruginosa detoxified mercury by means of volatilization from the M9 assay medium containing 20 ppm mercury. The highest removal rate was in the reaction with 1 ppm Hg concentration though the removal rate was quite good up to 8 ppm Hg in this medium and the removal rate decreased with increasing concentrations of NaCl. In medium amended with 100 ppm Cd, the concentration was reduced to 17.41 ppm by CH07 and to 19.19 ppm by GP06 in about 72 h. Thus, both CH07 and GP06 were capable of removing >70% Cd from growth medium. All three strains of MRB (CH07, S3 and GP13) precipitated Pb from the growth medium. In case of CH07 the concentration of Pb in medium amended with 100 ppm Pb reached as low as 1.8 ppm (>98% removal) in 96 h. As revealed by the SEM and EDS analyses, Pb was found to be entrapped in the exopolymeric substance (EPS). Isolate GP13 and S3 removed >87% Pb in the same period and precipitated it as lead sulfide. From this detailed study, it is clear that the MRB have cellular mechanisms to either immobilize the toxic heavy metals as in the case of Cd or precipitate them as in the case of Pb. Increases in the total protein content when exposed to heavy metals were recorded from all the isolates tested in presence of Hg or Cd which supports the view that the detoxification mechanisms involved expression of intracellular and/or extracellular proteins. Efficiency of these MRB in detoxifying the heavy metals like Hg, Cd, Pb and many other toxic substances provides a strong basis to hypothesize that these bacteria do use detoxification mechanisms, which are probably overlapping in their genetic nature sharing some common features. Precipitation of lead as lead sulfide has been an important finding probably either through extraneous formation or through intracellualer biochemical production of H₂S in aerobic condition from cysteine-containing enzymes.

Bacterial degradation of toxic xenobiotics studied during this study was very interesting in terms of efficiency and newer findings. Isolate CH07 was able to degrade **PCBs containing \geq 6 Chlorine atoms**. Among the different congeners of PCBs present in Clophen A-50, 14 chlorobiphenyls were degraded in varying percentages. Of these, one coplanar congener CB-126 and one sterically hindered CB-181were completely degraded within 40 h. This bacterium could detoxify the most toxic coplanar congeners, CB-77 and CB-126 in the Chlophen A-50 mixture. This unique process of degrading PCBs in less than two days was detected for the first time in a marine bacterium and a US patent (no. 6,544,773) was granted for this work. While the marine isolate CH07 was efficient in degrading/biotransforming PCBs in terms of chlorine atom removal, the **absence of** *bph***A and** *bph***C** in this isolate is quite puzzling and thus, demands further research to decipher the pathway if any involved in utilization of biphenyls through an alternative pathway.

The Pseudomonad, CH07 degraded TBT faster than *Alcaligenes faecalis*, GP15. At the end of the experiment, CH07 degraded nearly 54% of the initial TBT concentration whereas the amount was ca 34% by GP15. With organic enrichment, the total amount of TBT degraded was similar and, the degradation rate was also faster by both these strains. Efficiency of these MRB in **degradation of TBT** is quite promising and calling attention on their potential in bioremediation of some very deleterious environmental toxicants including toxic heavy metals as well as xenobiotics. Moreover, absence of the typical biphenyl dioxygenase in CH07 and its capability of degrading TBT, which is also known to be dioxygenase-mediated, makes one ponder to call in for investigations to checkout the presence of enzyme (s), enabling such bacteria for degrading a multitude of aromatic compounds.

Bioremediation of mercury from wastewater in a laboratory scale bioreactor over a period of one month has been demonstrated efficiently by this investigation. This process has been very effective, as high as 95%, recovery of the elemental mercury from the reactor. Moreover, CH07 was better adapted to higher salt concentration in the waste than the other isolate P. aeruginosa, Bro12 tested during this study. The cultures in the bioreactors were able to withstand different operational shocks including interruption in the nutrient supply, blockage of pipes due to bubble or salt precipitation etc. proving the sturdiness of the system (i.e., bacterial cultures) in efficient cleaning up of toxic mercury waste and their aiding its recovery, if desired. This process also ensures that the relatively less toxic gaseous mercury is not released into the atmosphere. Bioremediation of mercury-containing ASN-III medium to promote growth of mercury-sensitive *Phormidium* sp. was a successful demonstration of detoxification efficiency of these bacteria. Furthermore, bioremediation of agricultural soil prior to sowing of plants (in this study, a salt tolerant rice variety) further proves that these marine, mercury resistant bacteria are capable of performing in field conditions and treatment with these MRB prior to usage of soil for agricultural activity is very effective and quite promising.

If a single strain is constructed/trained to perform several metabolic activities, the efficiency and predictability of the process may be significantly enhanced. The marine pseudomonad strain, CH07 (NRRL B-30604) and many other strains isolated and identified during this study, prove their potential for bioremediation. These MRB form highly potential candidate-organisms for bioremediation of toxic wastes xenobiotics as well as many heavy metal/ or, persistent organic pollutants (POPs). They are also apparently useful in treating the contaminated sites in the open environments, although this study did not undertake any such field/site specific study. It can be

surmised that despite the alarming present scenario of chemical pollution, there is hope from these MRB possessing an array of armory for alleviating environmental health hazards.

Future Prospects:

During the course of this thesis many findings came up which were sometimes in agreement with earlier findings as well as contradictory. The hypothesis of this thesis has been well documented and supported with the results obtained although many questions have emerged in parallel, which warrant further studies and more in-depth investigations.

Enumeration of MRB on a regular basis should be carried out along with the regular microbiological analyses and this can become a regular parameter to realize the health-status of the environment. In principle, higher their abundance, more likely is the heavy metal contamination in that environment.

More detailed studies on heavy metal resistance form the marine environment should be carried out. Further investigation on mercury-resistance in marine bacteria may lead to new and better understanding of the existing concept. For instance, presence of non-*mer* mediated mercury volatilization in the marine bacteria might prove pivotal in acquiring more information on mercury-resistance. Also, further studies on the role of Na⁺ in transport of Hg across the cell membrane and role of Cl⁻ in determining the bioavailability are quite important in this regard. It is worth investigating if any of the enzymes and molecular mechanisms involved in mercury-resistance, also play a vital role in detoxification of other heavy metals.

The degradation pathway and the genes involved in the PCBs degradation by CH07 should be studied in details at molecular level. Since this is the first report of aerobic bacterial aerobic degradation of PCBs containing highly chlorinated (≥ 6) PCBs, more detailed and focused studies will help understand the mechanisms more succinctly.

The resistance of these marine MRB to several heavy metals enthuses to affirmatively recommend their potential to be exploited in bioremediation of mixed wastes. Further studies including on-site experiments will be useful in developing practical means for environmental clean up.

References

- 1. Abramowicz D. A. (1990). Aerobic and anaerobic biodegradation of PCBs: a review. Crit. Rev. Biotechnol. 3: 241-51.
- 2. Ahmed M. and Focht D. D. (1973). Degradation of polychlorinated biphenyls by two species of Achromobacter. Can. J. Microbiol. 19: 47-52.
- 3. Allen D. A., Austin B. and Colwell R. R. (1977). Antibiotic resistance patterns of metaltolerant bacteria isolated from an estuary. Antimicrob. Agents Chemother. 12: 545-47.
- Alzieu C. L. and Heral M. Ecotoxicological effects of organotin compounds on oyster culture. In : Ecotoxicological testing for the marine environment. (Personne et al. eds) vol. III. State University of Ghent, Ghent and Institute of Marine Science Research Bredence, Belgium. (1984). Pp. 187-96.
- 5. Alzieu C. L., Sanjuan J., Deltreil J. P. and Borel M. (1986). Tin contamination in Arcachon Bay-Effect on oyster shell anomalies. Mar. Poll. Bull. 17: 494-98.
- 6. Alzieu C. L., Sanjuan J., Michel P., Borel M. and Dreno J. P. (1989). Monitoring and assessment of butyltin in Atlantic coastal water. Mar. Poll. Bull. 20: 22-26.
- 7. Amy P.S. and Morita R. Y. (1983). Starvation-survival patterns of sixteen freshly isolated open-ocean bacteria. Appl. Environ. Microbiol. 45:1109-15.
- 8. Anon. (1998). Coastal Ocean Monitoring and Prediction System data (96-97). Tech. Rep. National Institute of Oceanography, Goa.
- 9. Anon. (1998). Marine Environmental Impact Assessment for Port Development at Poshitra Point, Okhamandal, Gujarat. Tech. Rep. National Institute of Oceanography, Goa.
- 10. Anon. (1999). Marine environmental impact assessment of proposed product pipeline between JNPT and IOCL's Vashi terminal. Tech. Rep. no. 8049 National Institute of Oceanography, Goa.
- 11. Anon. (1997). Current measurements for proposed all weather ports at Redi, Vijaydurg and Ratnagiri, Maharashtra. Tech. Rep. National Institute of Oceanography, Goa.
- 12. Anon. (1999). Deadly mettle: when wealth is not health. Down To Earth 7: 32-50.
- 13. Atlas R. M. (1991). Microbial hydrocarbon degradation. Bioremediation of oil spills. J. Chem. Technol. Biotechnol. 52: 149-56.
- 14. Bååth E., Diaz-Ravina M., Frostegård Å. and Campbell C. D. (1998). Effect of metal-rich sludge amendements on the soil microbial community. Appl. Environ. Microbiol. 64: 238-45.
- 15. Baldi F., Filippelli M. and Olson G. J. (1989). Biotransformation of mercury by bacteria isolated from a river collecting cinnabar mine waters. Microb. Ecol. 17: 263-74.
- 16. Baldwin D. R. and Marshall W. J. (1999). Heavy metal poisoning and its laboratory investigation. Ann. Clin. Biochem. 36: 267-300.
- 17. Bang S. W., Clark D. S. and Keasling J. D. (2000). Engineering hydrogen sulfide production and cadmium removal by expression of the thiosulfate reductase gene (phsABC) from Salmonella enterica serovar typhimurium in Escherichia coli. Appl. Environ. Microbiol. 66: 3939-44.
- 18. Barbieri P., Galassi G. and Galli E. (1989). Plasmid-encoded mercury resistance in a *Pseudomonas stutzeri* strain that degrades o-xylene. FEMS Microbiol. Ecol. 62: 375-84.

- 19. Barbieri P., Bestetti G., Reniero D. and Galli E. (1996). Mercury resistance in aromatic compound degrading *Pseudomonas* strains. FEMS Microbiol. Ecol. 20: 185-94.
- 20. Barkay T. and Olson B. H. (1986). Phenotypic and genotypic adaptation of aerobic heterotrophic sediment bacterial communities to mercury stress. Appl. Environ. Microbiol. 52: 403-06.
- 21. Barkay T. (1987). Adaptation of aquatic microbial communities to Hg²⁺ stress. Appl. Environ. Microbiol. 53: 2725-32.
- 22. Barkay T. and Pritchard P. H. (1988). Adaptation of Aquatic Microbial Communities to Pollutant Stress. Microbiol. Sci. 5(6): 165-69.
- 23. Barkay T., Liebert C. and Gillman M. (1989). Environmental significance of the potential for mer(Tn21)-mediated reduction of Hg²⁺ to Hg⁰ in natural waters. Appl. Environ. Microbiol. 55: 1196-202.
- 24. Barkay T., Turner R., Saouter E. and Hron J. (1992). Mercury biotransformation and their potential for bioremediation of mercury contamination. Biodegradation. 3: 147-59.
- 25. Barkay T., Gillman M. and Turner R. R. (1997). Effects of dissolved carbon and salinity on bioavailability of mercury. Appl. Environ. Microbiol. 63: 4267-71.
- 26. Barkay T. The mercury cycle. In Encyclopedia of microbiology, 2nd ed. Academic press, Inc., San Diego. (2000). Pp. 171-81.
- 27. Barkay T., Miller S. M. and Summers A. O. (2003). Bacterial mercury resistance from atoms to ecosystems. FEMS Microbiol. Rev. 27: 355-84.
- 28. Barreiro R., González R., Quintela M. and Ruiz J. M. (2001). Imposex, organotin bioaccumulation and sterility of female *Nassarius reticulatus* in polluted areas of NW Spain. Mar. Ecol. Prog. Ser. 218: 203-12.
- 29. Barug D. (1981). Microbial degradation of bis (tributyltin) oxide. Chemosphere. 10: 1145-54.
- 30. Bastidas C., Bone D. and García E. M. (1999a). Sedimentation rates and metal content of sediments in a Venezuelan coral reef. Mar. Poll. Bull. 38: 16-24.
- Bastidas C., Bone D. and García E. M. (1999b). Metal content on the Ref. Coral Porites asteroides: an evaluation of river influence and 35 years of chronology. Mar. Pollut. Bull. 38: 899-907.
- 32. Baumann L., Baumann P., Mandel M. and Allen R. D. (1972). Taxonomy of aerobic marine bacteria. J. Bacteriol. 110: 402-29.
- 33. Bedard D. L., Unterman R., Bopp L.H., Brennan M.J., Haberl M.L. and Johnson C. (1986). Rapid assay for screening and characterizing microorganisms for the ability to degrade polychlorinated biphenyls. Appl. Environ. Microbiol. 51: 761-68.
- Bedard D. L., Brannan Jr. and Michael J. (1987). Evidence for novel mechanisms of polychlorinated biphenyl metabolism in *Alcaligenes eutrophus* H850. Appl. Environ. Microbiol. 53: 1103-12.

- 35. Bedard D. and Quensen D. F. Microbial reductive dechlorination of polychlorinated biphenyls. In: Microbial transformation and degradation of toxic organic chemicals. Young L. and Cerniglia (Eds.). New York, Wiley-Liss. (1995). Pp. 127-216.
- 36. Benoit J. M., Gilmour C. C., Mason R. P. and Hayes A. (1999). Sulfide controls on mercury speciation and bioavailibility to methylating bacteria in sediment pore waters. Environ. Sci. Technol. 33: 951-58.
- Berkaw M, Sowers K. R. and May H. D. (1996). Anaerobic ortho dechlorination of Polychlorinated Biphenyls by Estuarine Sediments from Baltimore Harbor. Appl. Environ. Microbiol. 62: 2534-39.
- 38. Bizily S. J., Rugh C. L., Summers A. O. and Meagher R. B. (1999). Phytoremediation of methylmercury pollution: merB expression in *Arabidopsis thaliana* confers resistance to organomercurials. Proc. Nat. Acad. Sci. 96: 6808-6813.
- 39. Bizily S. P., Rugh C. L. and Meagher R. B. (2000). Phytodetoxification of hazardous organomercurials by genetically engineered plants. Nature Biotechnol. 18: 213-17.
- 40. Blair W. R., Olson G. J., Brinckman F. E. and Iverson W. P. (1982). Accumulation and fate of tri-n-butyltin cation in estuarine bacteria. Microb. Ecol. 8: 241-45.
- 41. Blake J. An approach to the regulation of bioremediation of polychlorinated biphenyls. In Hichee R. E. (Ed.), Bioremediation of chlorinated and polycyclic aromatic hydrocarbon compounds. Boca Ratón, Fl: Lewis Publishing Inc. (1994). Pp. 432-435.
- 42. Blunden S. J. and Chapman A. H. (1982). The environmental degradation of organotin compounds a review. Environ. Technol. Lett. 3: 267-72.
- 43. Bogdanova E. S., Mindlin S. Z., Pakrova E., Kocur M. and Rouch D. A. (1992). Mercuric reductase in environmental Gram-positive bacteria sensitive to mercury. FEMS Microbiol. Lett. 97: 95-100.
- 44. Bolton H. Jr. and Gorby Y. A. An overview of the bioremediation of inorganic contaminants. In Hinchee R.E., Means J. L. and Burris D. R. (ed.), Bioremediation of inorganics. Battelle Press, Columbus, Ohio. (1995). Pp. 1-6.
- 45. Boon J. P., Van Zantvroot M. B., Govaert M. J. M. A. and Duinker J. C. (1985). Organochlorines in benthic polychaetes (*Nephtys* spp.) and sediments from the southern North Sea. Identification of individual PCB components, Neth. J. Sea Res. 19: 93-109.
- 46. Boon J. P., Everaarts J. M., Kastoro W. W., Razak H., Sumanta I., Sumarno, Nelissen P.H., Stefels J. and Hillebrand M. T. J. (1989). Cyclic organochlorines in epibenthic organisms from coastal water around East Java, Neth. J. Sea Res. 23: 427-40.
- 47. Boudou A. and Ribeyre. Mercury in the food web: Accumulation and transfer mechanisms. *In* Sigel A. and Sigel H. (eds), metal ions in biological systems: mercury and its effects on environment and biology. Vol. 34. Marcel Dekker Inc., New York. (1997). Pp. 503-568
- 48. Boyle A.W., Silvin C. J., Hassett J. P., Nakas J. P. and Tanenbaum S.W. (1992). Bacterial PCB Biodegradation. Biodegradation 3: 285-98.

- 49. Bradford M. M. (1976). A rapid and sensitive method for the quantification of microgram quantities of protein utilizing the principle of protein-dye binding. Analyt. Biochem. 72: 248–54.
- 50. Bradley C. A., Kearns R. D., Wood P. P. and Black W. E. Degradation of polyhalogenated biphenyl compounds with white-rot fungus grown on sugar beet pulp. USP no. 5,583,041, December 10, 1996.
- Brim H., McFarlan S. C., Fredrickson J. K., Minton K., Zhai M., Wackett L. P. and Daly M. J. (2000). Engineering *Deinococcus radiodurans* for metal remediation in radioactive mixed waste environments. Nat. Biotechnol. 18: 85-90.
- 52. Brown N. L., Misra T. K., Winnie J. N., Schmidt A., Seiff M. and Silver S. (1986). The nucleotide of the mercuric resistance operon of plasmid R100 and transposon Tn501: further evidence for *mer* genes, which enhance the activity of the mercuric ion detoxification system. Mol. Gen. Genet. 202: 143-51.
- 53. Brown N. L., Camakaris J., Lee B. T., Williams T., Morby A. P., Parkhill J. and Rouch D. A. (1991). Bacterial resistance to mercury and copper. J. Cell. Biochem. 46: 106-14.
- 54. Bruce K. D., Osborn A. M., Pearson A. J., Strike P. and Ritchie D. A. (1995). Genetic diversity within mer genes directly amplified from communities in noncultivated soil and sediment bacteria. Mol. Ecol. 4: 605-12.
- 55. Brunke M., Deckwer W. D., Fritschmuth J. M., Horn H., Lunsdorf M., Rhode M., Rohricht M., Timmis K. N. and Weppen P. (1993). Microbial retention of mercury from waste systems in a laboratory column containing merA gene bacteria. FEMS Microbiol. Rev. 11: 45-52.
- 56. Cameron R. E. (1992). Guide to site and soil description for hazardous waste characterization. Vol. I: Metals. Environmental Protection Agency EPA/600/4-91/029.
- 57. Cànovas D., Cases I. and Lorenzo de V. (2003). Heavy metal tolerance and metal homeostasis in *Pseudomonas putida* as revealed by complete genome analysis. Env. Microb. 5: 1242-56.
- 58. Canstein V. H., Li Y., Timmis K. N., Deckwer W. D and Wagner-Döbler I. (1999) Removal of mercury from chloralkali electrolysis wastewater by a mercury-resistant *Pseudomonas putida* strain. Appl. Environ. Microbiol. 65: 5279-84.
- 59. Canstein H. V. F. (2000). Ph. D thesis "Mercury-reducing biofilms: insights in their activity and composition and application for bioremediation of mercury-polluted wastewaters. Technical University of Braunschweig, Germany.
- 60. Canstein V. H., Li Y. and Wagner-Döbler I. (2001). Long-term performance of bioreactors cleaning mercury-contaminated wastewater and their response to temperature and mercury stress and mechanical perturbation. Biotechnol Bioeng. 74: 212-19.
- Canstein V. H., Li Y., Leonhäuser J., Haase E., Felske A., Deckwer W. D. and Wagner-Döbler I. (2002). Spatially Oscillating Activity and Microbial Succession of Mercury-Reducing Biofilms in a Technical-Scale Bioremediation System. Appl. Environ. Microbiol. 68: 1938-46.
- 62. Champ M. A. (2000). A review of organotin regulatory strategies: pending actions, related costs and benefits. Sci. Tol. Environ. 258: 21-71.

- Champ M.A. Pollution prevention from ships and shipyards. In: proceedings of International Oceanology symposium-2001. Vol. 1, Miami, Florida, USA. (2001). Pp. 250.
- 64. Chang J. S., Hong J., Ogunseitan O. A. and Olson H. B. (1993). Interaction of mercuric ions with the bacterial growth medium and its effects in enzymatic reduction of mercury. Biotechnol. Prog. 9: 526-32.
- 65. Chang J. S. and Hong J. (1994). Biosorption of mercury by the inactivated cells of *Pseudomonas aeruginosa* PU21 (Rip64). Biotechnol. Bioeng. 44: 999-1006.
- 66. Chang J. S., Law. R. and Chang C. C. (1997). Biosorption of lead, copper and mercury by biomass of *Pseudomonas aeruginosa* PU21. Wat. Res. 31: 1651-58.
- 67. Chang J. S. and Law W. S. (1998). Development of microbial mercury detoxification process using a mercury-hyperesistant strain of *Pseudomonas aeruginoasa* PU21. Biotechnol. Bioeng. 57: 462-70.
- 68. Chen J. H., Lion L. W., Ghiorse W. C. and Shuler M. L. (1995). Mobilization of adsorbed cadmium and lead in aquifier material by bacterial extracellular polymers. Wat. Res. 29: 421-30.
- 69. Chen P. H., Wong C. K., Rappe C. and Nygren M. (1980). Polychlorinated biphenyls, dibenzofurans and quaterbiphenyls in toxic rice bran oil and in the blood and tissue of patients with PCB poisoning (Yu-Cheng) in Taiwan. Environ. Hlth. Perspect. 59: 59-65.
- 70. Chen S. and Wilson D. B. (1997a). Construction and characterization of Escherichia coli genetically engineered for bioremediation of Hg (2⁺) contaminated environments. Appl. Environ. Microbiol. 63: 2442-45.
- 71. Chen S. and Wilson D. B. (1997b). Genetic engineering of bacteria and their potential for Hg²⁺ bioremediation. Biodegradation. 8: 97-103.
- 72. Chen S., Kim E., Shuler M. L. and Wilson D. B. (1998). Hg²⁺ removal by genetically engineered *Escherichia coli* in a hollow fiber bioreactor. Biotechnol. Prog. 14: 667-71.
- 73. Chen S. H. and Aitken M. D. (1999). Salicylate stimulates the degradation of high molecular weight polycyclic aromatic hydrocarbons by *Pseudomonas saccharophila*. Env. Sci. Technol. 33: 435-39.
- 74. Churchill P. F., Dudley R. J. and Churchill S.A. (1995). Surfactant-Enhanced Bioremediation. Waste Management. 15: 371-77.
- 75. Colwell R. R., Brayton P. R., Grimes D. J., Roszak D. B., Huq S. A. and Palmer L. M. (1985) Viable, but non-culturable *Vibrio cholerae* and related pathogens in the environment: implications for release of genetically engineered microorganisms. Biotechnology. 3: 817-20.
- 76. Compeau G. and Bartha R. (1984). Methylation and demethylation of mercury under controlled redox, pH and salinity conditions. Appl. Environ. Microbiol. 48: 1203-07.
- 77. Condee C. W. and Summers A. O. (1992). A mer-lux transcriptional fusion for real-time examination of in vivo gene expression kinetics and promoter response to altered superhelicity. J. Bacteriol. 174: 8094-101.

- Cooney J. J. (1988). Microbial transformation of tin and tin compounds. J. Ind. Microbiol. 3: 195-204.
- 79. Conney J. J. and Wuertz S. (1989) Toxic effects of tin compounds on microorganisms. J. Ind. Microbiol. 4: 375-402.
- 80. Cooney J. J., de Rome I., Laurence O. S. and Gadd G. M. (1989). Effect of organotin and organolead compounds on yeast. J. Ind. Microbiol. 4: 279-88.
- 81. Cooney J. J. (1995). Organotin compounds and aquatic bacteria- a review. Helgoland Meeresunter. 49: 663-77.
- 82. Cowan S. T and Steel K. J. Manual for the identification of medical bacteria. (eds.), Cambridge University Press. London, New York, Melbourne. 2nd edn., (1974). Pp. 238.
- 83. D'Itri F. M. The biomethylation and cycling of selected metals and metalloids in aquatic sediments. *In*: Sediments: Chemistry and Toxicity of In-Place Pollutants (R. Baudo, J. Giesy and H. Muntau, eds), Lewis, Ann Arbor, MI. (1990).
- 84. Daly M. J. (2000). Engineering radiation-resistant bacteria for environmental biotechnology. Curr. Opin. Biotechnol. 11: 280-85.
- 85. De J., Ramaiah N., Mesquita A. and Verlekar X. N. (2003). Tolerance to various toxicants by marine bacteria highly resistant to mercury. Mar. Biotechnol. 5: 185-93.
- 86. De Mora S. J. Tributyltin-case study of an environmental contaminant. Cambridge University Press. Cambridge, UK. (1996).
- 87. De Rore H., Top E., Houwen F., Mergcay M. and Verstraete W. (1994). Evolution of heavy metal-resistant transconjugants in a soil environment with a concomitant selective pressure. FEMS Microbiol. Ecol. 14: 263-73.
- 88. de Souza M. P., Huang C. P., Chee N. and Terry N. (1999). Rhizosphere bacteria enhance the accumulation of selenium and mercury in wetland plants. Planta. 209: 259-63.
- 89. Deckwer W. D., Becker F. U., Ledakowicz S. and Wagner-Döbler I. (2004). Microbial removal of ionic mercury in a three-phase fluidized bioreactor. Env. Sci. Technol. 38: 1858-65.
- 90. Deng X. and Wilson D. B. (2001). Bioaccumulation of mercury from wastewater by genetically engineered *Escherichia coli*. Appl. Microbiol. Biotechnol. 56: 276-79.
- 91. Diels L., Dong Q., Van der Lelie D., Baeyens W. and Mergeay A. (1995). The *czc* operon of *Alcaligenes eutrophus* CH34: from resistance mechanism to the removal of heavy metals. J. Ind. Microbiol. 14: 142-53.
- 92. Doelman P., Jansen E., Michels M. and Van Til. M. (1994). Effects of heavy metals in soil on microbial diversity and activity as shown by the sensitivity-resistance index, an ecologically relevant parameter. Biol. Fertil. Soil. 17: 177-84.
- 93. Dong D., Nelson Y. M., Lion L. W., Shuler M. L. and Ghiorse W. M. (2000). Adsorption of Pb and Cd onto metal oxides and organic material in natural surface coatings as determined by selective extractions: new evidence for the importance of Mn and Fe oxides. Wat. Res. 34: 427-36.
- 94. Dowson P. H., Bubb J. M., Williams T. P. and Lester J. N. (1993). Degradation of tributyltin in freshwater and estuarine marine sediments. Wat. Sci. Technol. 28: 133-37.

- 95. Dua M., Singh A., Sethunathan N. and Johri A. K. (2002). Biotechnology and bioremediation: success and limitations. Appl. Microbiol. Biotechnol. 59:143-52.
- 96. Dubey S. K. and Roy U. (2003). Biodegradation of tributyltins (organotins) by marine bacteria. Appl. Organometal. Chem. 17: 3-8.
- 97. Dunn B. P., Black, J. J. and Maccubbin A. (1987) ³²P-postlabelling analysis of aromatic DNA adducts in fish from polluted water. Cancer Res. 47: 6543-48.
- 98. EPA, U. S. E. P. A. (1998). Analysis of current mercury waste generation and treatment. http://www.epa.gov/.
- 99. Essa A. M., Macaskie L. E. and Brown N. L. (2002). Mechanisms of mercury bioremediation. Biochem. Soc. Trans. 30: 672-74.
- 100. Evans S. M. (1999). TBT or no TBT? That is the question. Biofouling. 14: 117-29.
- 101. Everaarts, J. M., Sleiderink, Hedwig M., Besten, Pieter J. Den, Halbrook, Richard S., and Shugart Lee R. (1994) Molecular responses as indicators of marine Pollution: DNA damage and enzyme induction in *Limamnda limanda* and *Asterias rubens*. Environ. Hlth. Perspect. 102: 37-43.
- 102. Fassett D. W. Chapter 4: Cadmium. In: Lee DHK (ed). Metallic contaminants and human health. 1st ed. New York: Academic Press; 1972. Pp. 97-124.
- 103. Fein J. B. and Delea D. (1999). Experimental study of the effect of EDTA on Cd adsorption by *Bacillus subtilis*: a test of the chemical equilibrium approach. Chem. Geol. 161: 375-83.
- 104. Fitzgerald W. F. and Mason R. P. Biogeochemical cycling of mercury in the marine environment, In A. Sigel and H. Sigel (eds), Mercury and its effect on environmental biology, 1st ed. Mercel Dekker, Inc., New York, N. Y. (1997). Pp. 53-111.
- 105. Fitzgerald W. F., Vandal G. M., Tolfhus K. R., Lamborg C. H. and Langer C. S. (2000). Mercury emissions and cycling in the coastal zone. J. Environ. Sci. 12: 92-101.
- 106. Förstner U., Wittmann G. T. W. Metal pollution in the aquatic environment. Springer Verlag Berlin Heidelberg New York. (1979). Pp. 486.
- 107. Frischmuth A., Weppen P. and Deckwer W. D. (1991).Quecksilberentfernung aus wässerigen Medien durch aktive mikrobielle Prozesse. Bioengineering. 7: 38.
- 108. Fry J. C., Gadd G. M., Herbert R. A., Jones C. W. and Watson C. I. A (eds.). Microbial control of pollution. Proceeding of the 48th symposium of the society for general microbiology held at the University of Cardiff, March 1992. Cambridge University Press. (1992)..
- 109. Fukagawa T. and Suzuki S. (1993). Cloning of gene responsible for tributyltin chloride (TBTCl) resistance in TBTCl-resistant marine bacterium *Alteromonus* sp. M-1. Biochem. Biophys. Res. Commun. 194: 733-40.
- 110. Fukagawa T., Konno S., Takama K. and Suzuki S. (1994) Occurrence of tributyltin (TBT) and methylmercury tolerant bacteria to which TBT was added. J. Mar. Biotechnol. 1: 211-14.

- 111. Fukuda Y., Ushijima K., Sakamoto M. and Futatsuka M. (1999). An analysis of subjective complaints in a population living in a methylmercury-polluted area. Env. Res. 81: 100-07.
- 112. Furukawa K., Tomizuka N., and Kamibayashi A. (1979). Effect of chlorine substitution on the bacterial metabolism of various polychlorinated biphenyls. Appl. Environ. Microbiol. 38:301-10
- 113. Furukawa K. (2000). Biochemical and genetic bases of microbial degradation of polychlorinated biphenyls (PCBs). J. Gen. Appl. Microbiol. 46: 283-96.
- 114. Gadd G. M. (1990). Heavy metal accumulation by bacteria and other microorganisms. Experientia. 46: 834-40.
- 115. Gadd G. M., Gray D. J. and Newby P. J. (1990). Role of melanin in fungal biosorption of tributyltin chloride. Appl. Microbiol. Biotechnol. 34: 116-69.
- 116. Gadd G. M. and White C. (1993). Microbial treatment of metal pollution- a working biotechnology? Trends Biotechnol. 11: 353-59.
- 117. Gadd G. M. (2000). Microbial interactions with tributyltin compounds: detoxification, accumulation and environmental fate. Sci. Tot. Environ. 258: 119-27.
- 118. Garcia E. M., Orta M. and Suarez P. (1999). Toxicity assays and bioconcentration of mercury in bacteria selected from marine environments. Bull. Environ. Contam. Toxicol. 62: 79-86.
- 119. Gerlach A. S. Marine Pollution. Diagnosis and Therapy. Springer-Verlag Berlin Heidelberg New York. (1981). Pp. 218.
- 120. Gibson D. T. and Sayler G.S. Scientific Foundations of Bioremediation: Current Status and Future Needs. American Academy of Microbiology, Washington, D.C., USA. (1992).
- 121. Gilmour C. C., Henry E. A. and Mitchell R. (1992). Sulfate stimulation of mercury methylation in freshwater sediments. Env. Sci. Technol. 26: 2281-87.
- 122. Goldwater L. J. and Clarkson T. W. Chapter 2: Mercury. In: Lee DHK (ed). Metallic contaminants and human health. 1st ed. New York: Academic Press. (1972). Pp.17-55.
- 123. Goyer R. A. and Chisholm J. J. Chapter 3: Lead. In: Lee DHK (ed). Metallic contaminants and human health. 1st ed. New York: Academic Press. (1972). Pp.57-95.
- 124. Griffin H. G., Foster T. J., Silver S. and Misra T. K. (1987). Cloning and DNA sequence of the mercuric and organomercurial resistance determinates of plasmid pDU1358. Proc. Natl. Acad. Sci. USA. 84: 3112-16.
- 125. Hamlett N. V., Landale E. C., Davis B. H. and Summers A. O. (1992). Roles of the Tn21 merT, merP and merC gene products in mercury resistance and mercury binding. J. Bacteriol. 174: 6377-85.
- 126. Harithsa S., Kerkar S. and Bharathi P. A. L. (2002). Mercury and lead resistance in hypersaline sulfate-reducing bacteria. Mar. Poll. Bull. 44: 726-32.

- 127. Hassan M. E. T., Van der Lelie D., Springael D., Römling N., Ahmed N. and Mergeay M. (1999). Identification of a gene cluster, czr, involved in cadmium and zinc resistance in *Pseudomonas aeruginosa*. Gene. 238: 417-25.
- 128. Heaton A. C., Rugh C. L., Kim T., Wang N. J. and Meagher R. B. (2003). Toward detoxifying mercury-polluted aquatic sediments with rice genetically engineered for mercury resistance. Environ. Toxicol. Chem. 22: 2940-47.
- 129. Hickey W. J. and Focht D. D. (1990). Degradation of monohalogenated, dihalogenated and trihalogenaed benzoic acids by *Pseudomonas aeruginosa* jb2. Appl. Environ. Mirobiol. 56: 3842-50.
- 130. Hideomi N., Tomoaki I., Yasunaga S., Isamu K. and Susumu M. (1977). Frequency of heavy- metal resistance in bacteria from inpatients in Japan. Nature. 266: 165-67.
- 131. Hiroki M. (1994). Populations of Cd-tolerant microorganisms in soil polluted with heavy metals. Soil Sci. Plant. Nutr. 40: 515-24.
- Hobman J. L. and Brown N. L. Bacterial Mercury-Resistance Genes. In H. Sigel and A. Sigel (eds.). Metal Ions in Biological Systems Vol. 34. Marcel Dekker Inc. New York, USA. (1997). Pp. 527-568.
- 133. Hobman J., Kholodii G., Nikiforov V., Ritchie D. A., Strike P. and Yurieva O. (1994). The sequence of the *mer* operon of p327/419, 330 and 05. Gene. 277: 73-78.
- 134. Horn J. M., Brunke M., Deckware W. D. and Timmis K. N. (1994). *Pseudomonas putida* strains which constitutively overexpress mercury resistance for biodetoxification of organomercurial pollutants. Appl. Environ. Microbiol. 60: 357-62.
- 135. Horvat M., Covelli S., Faganeli J., Logar M., MandiŽ V., Rajar R., Žirca A. and DuŽan Ž. (1999). Mercury in contaminated environments; a case study: the Gulf of Trieste. Sci. Tot. Env. 237/238: 43-56.
- Hu H. Chapter 397: Heavy metal poisoning. In: Fauci AS, Braunwald E, Isselbacher K. J., Wilson J. D., Martin J. B., Kasper D. L., Hauser S. L. and Longo D. L. (eds). Harrison's principles of Internal medicine. 14th ed. New York: McGraw-Hill. (1998). Pp. 2564-69.
- 137. Hughes M. N. and Poole R. K. Metals and Micro-organisms. Chapman and Hall, London. (1989). Pp. 303-58.
- 138. Hütter L. A. (1978). Quecksilbergefährdung durch Leuchtstofflampen? Naturwiss Rundsch 3a: 17-19.
- 139. Inoue C., Sugawara K. and Kusano T. and Kitagawa (1989). Nucleotide sequence if the *Thiobacillus ferrooxidans* chromosomal gene encoding mercuric reductase. Gene. 84: 47-54.
- 140. Inoue C., Sugawara K. and Kusano T. (1991). The *mer*R regulatory gene in *Thiobacillus ferrooxidans* is spaced apart from the mer structural genes. Mol. Microbiol. 5: 2707-18.
- 141. Inoue H., Takimura O., Fuse H., Murakami K and Yamaoka Y. (2000). Degradation of triphenyltin by a fluorescent Pseudomonad. App. Environ. Microbiol. 66: 3492-98.

- 142. Inouhe M., Sumiyoshi M., Tohoyama H. and Joho M. (1996). Resistance to cadmium ions and formation of a cadmium-binding complex in various wild-type yeasts. Plant Cell Physiol. 37: 341-46.
- 143. Iohara K., Iiyama R., Nakamura K., Silver S., Sakai M., Takeshita M. and Furukawa K. (2001). The mer operon of a mercury-resistant *Pseudoalteromonas haloplanktis* strain isolated from Minamata Bay, Japan. Appl. Microbiol. Technol. 56: 736-41.
- 144. Izaki K. (1981). Enzymatic reduction of mercurous and mercuric ions in *Bacillus cereus*. Can. J. Microbiol. 27: 192-97.
- 145. Ji G. and Silver S. (1995). Bacterial resistance mechanisms for heavy metals of environmental concern. J. Ind. Microbiol. 14: 61-75.
- 146. Ka J. O., Holben W. E. and Tiedje J. M. (1994). Genetic and phenotypic diversity of 2, 4-dichlorophenoxyacetic acid (2, 4-D)-degrading bacteria isolated from 2, 4-D treated field soils. Appl. Environ. Microbiol. 60: 1106-15.
- 147. Kaladharan P., Pillai V. K., Nandakumar A. and Krishnakumar P. K. (1999). Mercury in seawater along the west coast of India. Ind. J. Mar. Sci. 28: 338-40.
- 148. Kaushik B. D. and Goyal S. K. (1993). Laboratory Manual, Seventh Training Course in Blue Green Algae, IARI Publications.
- Kawai S., Kurokawa Y., Harino H. and Fukushima M. (1998). Degradation of tributyltin by a bacterial strain isolated from polluted river water. Environ. Poll. 102: 259-63.
- 150. Kennedy V. S. The estuary as a filter. In Proceedings of the Seventh Biannual International Estuarine Research Conference, Virginia Beach, Va. Academic Press, Inc., New York. (1984).
- 151. Khalil Z. (1997) Toxicological response of a cyanobacterium, *Phormidium fragile* to mercury. Water air soil Poll. 98: 179-85.
- 152. Kholodii G. Y., Yurieva O. V., Lomovskaya O. L., Gorlenko Z. M., Mindlin S. Z. and Nikiforov V. G. (1993). Tn5053, a mercury resistance transposon with integron ends. J. Mol. Biol. 230: 1103-07.
- 153. Kim K. S., Pai H. S., Lee S. Y. and Ryu, D. D. Y. (1990). Effect of intercalating dyes on the production of antibiotics by *Microspora rosaria* and *Micromonospora purpurea*. Enzyme. Microbiol. Technol. 12 (8): 564-70.
- 154. Kim C. S., Brown Jr. G. E. and Rytuba J. J. (2001). Characterization and speciation of mercury-bearing mine wastes using X-ray absorption spectroscopy. Sci Tot. Environ. 261: 157-68.
- 155. Kimbara K., Shimura M., Hatta T., Kiyohara H. Method for degrading polychlorinated biphenyls and novel microorganism. USP no. 5,989,896; November 23, 1999.
- 156. Kimbrough R. D. (1987). Human health effects of PCBs and PBBs. Ann. Rev. Pharmacol. Toxicol. 27: 87-111.

- 157. Kiran Y. V and Anil A. C. (1999). Record of imposex in *Cronia konkanesis* (Gastropoda, Muricidae) from Indian waters. Mar. Environ. Res. 48:123-30.
- 158. Kiyono M., Omura T., Inuzuka M., Fujimori P. and Pan-Hou H. (1997). Nucleotide sequence and expression of the organomercurial-resistance determinants from a *Pseudomonas* K-62 plasmid pMR26. Gene. 189: 151-57.
- 159. Kiyono M. and Pan-Hou H. (1999). The merG Gene Product is Involved in Phenylmercury Resistance in *Pseudomonas* Strain K-62. J. Bacteriol. 181: 726-30.
- 160. Kiyono M., Uno Y., Omura T. and Pan-Hou H. (2000). Involvement of merB in the expression of the pMR26 mer operon induced by organomercurials. J. Hlth. Sci. 46: 142-45.
- Kiyono M., Omura H., Omura T., Murata S. and Hou-Pan H. (2003). Removal of inorganic and organic mercurials by immobilized bacteria having mer-ppk fusion plasmids. Appl. Microbiol. Biotechnol. 62: 274-78.
- 162. Klaassen C. D., Amdur M. O., Doull J. Toxicology. III ed. The basic science of poisons. Macmillan, New York, NY. (1986).
- 163. Knight C., Kaiser J., Lalor G. C., Robotham H. and Witter J. V. (1997). Heavy metals in surface water and stream sediments in Jamiaca. Environ. Geochem. Hlth. 19: 63-66.
- 164. Kogure K., Simidu U, Taga N. and Colwell R. R. (1987). Correlation of direct viable counts with heterotrophic activity for marine bacteria. Appl. Environ. Microbiol. 53: 2332-37.
- 165. Komura I. and Izaki K. (1971). Mechanism of mercuric chloride resistance in microorganisms. I. Vaporization of a mercury compound from mercuric chloride by multiple drug resistance strain of *Escherichia coli*. J. Biochem. 70: 885-93.
- Kovalic W. Perspective on risks of soil pollution and experience with innovative remediation technologies. Strategies 2000 Proc. World Congr. Chem. Eng. 4th ed. (1991). Pp. 281-95.
- 167. Krieg N. R., Holt G. J., (Eds. in chief). Bergey's manual of systematic bacteriology. Vol.1, Williams and Wilkins, Baltimore. (1984). Pp. 964.
- 168. Kusano T., Ji G., Inoue C. and Silver S. (1990). Constitutive synthesis of a transport function encoded by the *Thiobacillus ferrooxidans mer*C gene cloned in *Escherichia coli*. J. Bacteriol. 172: 2688-92.
- 169. Lacerda L. D. and Pfeiffer W. C. (1992). Mercury from gold mining in Amazon environment an overview. Quimica Nova. 55: 238-94.
- 170. Laemmli U. K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophase T4. Nature. 227: 680-85.
- 171. Langford N. J. and Ferner R. E. (1999). Toxicity of mercury. J. Hum. Hypertension. 13: 651-56.
- 172. Langley S. and Beveridge T. J. (1999). Effect of o-side-chain-lipopolysaccharide chemistry on metal binding. Appl. Environ. Microbiol. 65: 489-98.

- 173. Laughlin R.B., French W. and Guard H. E. (1986). Accumulation of bis (trobutyltin) oxide by the marine mussel, *Mytilus edulis*. Environ. Sci. Technol. 20: 884-90.
- 174. Lebeau T., Bagot D., Jézéquel K. and Fabre B. (2002). Cadmium bioabsorption by free and immobilized microorganisms cultivated in a liquid soil extract medium: effects of Cd, pH and techniques of culture. Sci. Tot. Environ. 291: 73-83.
- 175. Ledin M. (2001). Accumulation of metals by microorganisms- processes and importance for soil systems. Earth-science Rev. 51: 1-31.
- 176. Lee S-W., Glickmann E. and Cooksey A. (2001). Chromosomal locus for cadmium resistance in *Pseudomonas putida* consisting of cadmium-transporting ATPase and a MerR family response regulator. Appl. Environ. Microbiol. 67: 1437-44.
- 177. Liebert C. and Barkay T. (1988). A direct viable counting method for measuring tolerance of aquatic microbial communities to Hg²⁺. Can. J. Microbiol. 34: 1090-95.
- 178. Liebert C. A., Wireman J., Smith T. and Summers A. O. (1997). Phylogeny of mercury resistance (mer) operons from gram-negative bacteria isolated from the fecal flora of primates. Appl. Environ. Micobiol. 63: 1066-76.
- 179. Liebert C. A., Hall R. M. and Summers A. O. (1999). Transposon Tn21, flagship of the floating genome. Microbiol. Mol. Biol. Rev. 63: 507-22.
- 180. Livingstone C. Chapter 16: Acute metallic poisonings. In: Matthew J, Lawson A.
 A. H. Treatment of common acute poisonings. 4th ed. Edinburgh. (1979). Pp.129-38.
- 181. Lowry O. H., Rosebrough N. J., Farr A. L. and Randall R. J. (1951). Protein measurement with the Folin-phenol reagents. J. Biol. Chem. 193: 265-75.
- 182. Lu C. M., Chau C. W. and Zhang J. H. (2000). Acute toxicity of excess mercury on the photosynthetic performance of cyanobacterium, *S. platensis* assessment by chlorophyll fluorescence analysis. Chemosphere. 41: 191-96.
- 183. Lund P. A., Ford S. J. and Brown N. L. (1986). Transcriptional regulation of the mercury-resistance genes of transposon Tn501. J. Gen. Microbiol. 132: 465-80.
- 184. Lund P. A. and Brown N. L. (1987). Role of the merT and merP gene products of transposon Tn501 in the induction and expression of resistance to mercuric ions, Gene. 52:207-14.
- 185. Macalady J. L., Mack E. E., Nelson D. C. and Scow K. M. (2000). Sediment microbial community structure and mercury methylation in mercury-polluted Clear Lake, California. Appl. Environ. Microbiol. 66: 1479-88.
- 186. Macdonald R. W., Barrie L. A., Bidleman T. F., Diamond M. L., Gregor D. J. Semkin R. G., Strachan W. M., Li Y. F., Wania F., Alaee M., Alexeva L. B., Backus S. M., Bailey R., Bewers J. M., Gobeil C., Halsall C. J., Harner T., Hoff J. T., Jantunen L. M., Lockhart W. L., Mackay D., Muir D. C., Pudykiewicz J., Reimer K. J., Smit J. N. and Stern G. A. (2000). Contaminants in the Canadian Arctic: 5 years of progress in understanding sources, occurrences and pathways. Sci. Tot. Env. 254: 93-234.
- 187. MacFaddin F. J. Biochemical tests for identification of medical bacteria. Williams and Wilkins (eds.). Baltimore, 2nd edition. (1980). Pp. 527.

- 188. Magalhães de A. E. M. and Tubino M. (1995). A possible path for mercury in biological systems: the oxidation of metallic mercury by molecular oxygen in aqueous solutions. Sci. Tot. Environ. 170: 229-39.
- 189. Maguire R. J., Tkacz R. J., Chau Y. K., Bengert G. A. and Wong P. T. S. (1986). Occurrence of organotin compounds in water and sediment in Canada. Chemosphere. 15: 253-74.
- 190. Mahtani S. and Mavinkurve S. (1979). Microbial purification of longifolene-Asesquiterpene. J. Ferment. Technol. 57: 529-33.
- 191. Mason R. P., Fitzgerald W. F. and Morel M. M. (1994). The biogeochemical cycling of elemental mercury: anthropogenic influences. Geochim. Cosmochim. Acta. 58: 3191-98.
- 192. Matsuda K., Kobayashi E., Okubo Y., Suwazono Y., Kido T., Nishijo M., Nakagawa H. and Nogawa K. (2003). Total cadmium intake and mortality among residents in the Jinzu River Basin, Japan. Arch. Environ. Hlth. 58: 218-22.
- 193. Matthias C. L., Bellama J. M., Olson G. J. and Brinckman F. E. (1986). Comprehensive method for determination of aquatic butyltin and butylmethyltin species at ultratrace levels using simultaneous hybridization extraction with gas chromatography flame photometric detection. Env. Sci. Technol. 20: 609-15.
- 194. McArthur J. V. and Tuckfield R. C. (2000). Spatial patterns in among stream bacteria: effects of industrial pollution. Appl. Environ. Microbiol. 66: 3722-26.
- 195. McGraph R. (1995). Managing sediment remediation costs. Pollution Engineering. 27, 34-35.
- 196. Meinhardt F., Schaffrath R. and Larsen M. (1997). Microbial linear plasmids. Appl. Microbiol. Biotechnol. 47: 329-36.
- 197. Miller S. M., Ballou D. P., Massey V., Williams C. H. Jr. and Walsh C. T. (1986). Two-electron reduced mercuric reductase binds Hg (II) to the active site of dithiol but does not catalyze Hg (II) reduction. J. Biol. Chem. 261: 8081-84.
- 198. Minchin D., Bauer B., Oehlmann J. and Duggan C. B. (1997). Biological indicator used to map organotin contamination from a fishing port. Killeybegs, Ireland. Mar. Poll. Bull. 34: 235-43.
- 199. Misra T. K., Brown N. L., Fritzinger D. C., Pridmore R. D., Barnes W. M., Haberstroh L. and Silver S. (1984). The mercuric-ion resistance operon of plasmid R100 and transposon Tn501: the beginning of the operon including the regulatory region and the first two structural genes. Proc. Natl. Acad. Sci. USA. 81: 5975-79.
- 200. Misra T. K. (1992). Bacterial resistance to inorganic mercury salts and organomercurials. Plasmid. 27: 4-16.
- 201. Morby A. P., Hobman J. L. and Brown N. L. (1995). The role of cysteine residues in the transport of mercuric ions by the Tn501 MerT and MerP mercury-resistance proteins. Mol. Microbiol. 17: 25-35.
- 202. Moreira J. C. and Piveta F. (1997). Human and environmental contamination by mercury from industrial use in Brazil. Water Air Soil Pollut. 97: 241-46.

- 203. Morel F. M. M. and Hering J. G. Principles and Applications of Aquatic Chemistry. Wiley-Interscience, New York. (1993). Pp. 588.
- 204. Morel F. M. M, Kraepiel A. M. L. and Amyot M. (1998). The chemical cycle and bioaccumulation of mercury. Ann. Rev. Ecol. Syst. 29: 543-66.
- 205. Müller A. K., Rasmussen L. D. and Sorensen S. J. (2001a). Adaptation of the bacterial community to mercury contamination. FEMS Microbiol. Lett. 16: 49-53.
- 206. Müller K. A., Westergaard K. and Christensen S. (2001b). The effect of long-term mercury pollution on the soil microbial community. FEMS Microbiol. Ecol. 36: 11-19.
- 207. Mulligan C. N., Yong R. N. and Gibbs B. F. (2001). Remediation technologies for metal-contaminated soils and groundwater: an evaluation. Engineering Geol. 60:193-207.
- 208. Murthy S. D. S. and Mohanty P., (1993). Mercury ions inhibit photosynthetic electron transport at multiple sites in the cyanobacterium *Synechococcus* 6301. J. Biosci. 18: 355-60.
- 209. Naganuma T. (1996). Differential enumeration of intact and damaged marine planktonic bacteria based on cell membrane integrity. J. Aquat. Ecosys. Hlth. 5: 217-22.
- 210. Nagayama L., Kuratsune M. and Masuda Y. (1976). Determination of chlorinated dibenzofurans in Kanechlors and "Yosho oil". Bull. Environ. Contam. Toxicol. 15: 9-13.
- 211. Nahida B., Majed A. and Preston M. R. (2000). An assessment of the total mercury and methyl mercury content of zooplankton and fish tissue collected from Kuwait territorial waters. Mar. Poll. Bul. 40: 298-307.
- 212. Nakamura K., Fujisaki T. and Tamashiro H. (1986). Characteristics of Hgresistant bacteria isolated from Mina Mata Bay sediment. Environ. Res. 40: 58-67.
- Nakamura K. and Nakahara H. (1988a). Simplified x-ray film method for detection of bacterial volatilization of mercury chloride by *Escherichia coli*. Appl. Env. Microbiol. 54: 2871-73.
- 214. Nakamura K., Fujisaki T. and Shibata Y. (1988b). Mercury-resistant bacteria in the sediment of Minamata Bay. Nippon Suisan Gakkaishi. 54: 1359-63.
- 215. Nakamura K., Mineshi S., Uchiyama H. and Yagi O. (1990). Organomercurial volatilizing bacteria in the mercury polluted sediment of Minamata Bay, Japan. Appl. Environ. Microbiol. 56: 304-05.
- 216. Nascimento A. M. A. and Chartone-Souza E. (2003). Operon mer: bacterial resistance to mercury and potential for bioremediation of contaminated environments. Genet. Mol. Res. 2: 92-101.
- 217. Nierenberg D. W., Nordgren R. E., Chang M. B., Siegler R. W., Blayney M. B., Hochberg F., Toribara T. Y., Cernichiari E. and Clarkson T. (1998). Delayed cerebellar disease and death after accidental exposure to dimethylmercury. New England J. Med. 338: 1672-76.
- 218. Nies D. H., Nies A., Chu L. and Silver S. (1989). Expression and nucleotide sequence of a plasmid-determined divalent cation efflux system from *Alcaligenes euthophus*. Proc. Natl. Acad. Sci. USA. 86: 7351-55.

- 219. Nies D. H. (1995). The cobalt, zinc and cadmium efflux system CzcABC from *Alcaligenes eutrophus* functions as a cation-proton antiporter in Escherichia coli. J. Bacteriol. 177: 2707-12.
- 220. Nies D. H. and Silver S. (1995). Ion efflux system involved in bacterial metal resistances. J. Indust. Microbiol. 14: 186-99.
- 221. Nies D. H. (1999). "Microbial heavy-metal resistance". Appl. Microbiol. Biotechnol. 51: 730-50.
- 222. Nogawa K. and Ishizaki A. (1979). A comparison between cadmium in rice and renal effects among inhabitants of the Jinzu River Basin. Environ. Res. 18: 410-20.
- 223. Nriagu J. O. (1990). Global metal pollution. Poisoning the biosphere? Environment. 32: 7-33.
- 224. Nucifora G., Chu L., Silver S. and Misra T. K. (1989). Mercury operon regulation by the merR gene of the organomercurial resistance system of plasmid pDU1358. J. Bacteriol. 171: 4241-47.
- 225. Ogunseitan O. A. (2002). Episodic bioavailibility of environmental mercury: implications for biotechnological control of mercury pollution. Afr. J Biotechnol. 1: 1-9.
- 226. Okino S., Iwasaki K., Yagi O. and Tanaka H. (2002). Removal of mercuric chloride by a genetically engineered mercury-volatilizing bacterium *Pseudomonas putida* PpY101/pSR134. Bull. Environ. Contam. Toxicol. 68: 712-19.
- 227. Oliver J. D. (1982). Taxonomic scheme for the identification of marine bacteria Deep-Sea Res. 29 (A): 795-98.
- 228. Oliver J. D., Hite F., McDougald D., Andon N. L. and Simpson L. M. (1995) Entry into, and resuscitation from, the viable but nonculturable state by *Vibrio vulnificus* in an estuarine environment. Appl. Environ. Microbiol. 61: 2624-30.
- 229. Oliver J. and Solano B. (1998). Mercuy in environmental samples from a water body contaminated by gold mining in Colombia, South America. Sci. Tol. Environ. 217: 83-89.
- 230. Olson B., Barkay T. B., Nies D., Bellama J. M. and Colwell R. R. (1999). Plasmid mediation of mercury volatilization and methylation by estuarine bacteria. J. Ind. Microbiol. Biotechnol. 22: 418-27.
- 231. Osborn A. M., Bruce K. D., Strike P. and Ritchie D. A. (1993). Polymerase chain reaction-restriction fragment length polymorphism analysis shows divergence among mer determinants from gram-negative soil bacteria indistinguishable by DNA-DNA hybridization. Appl. Environ. Microbiol. 59: 4024-30.
- 232. Osborn A. M., Bruce K. D., Strike P. and Ritchie D. A. (1997). Distribution, diversity and evolution of the bacterial mercury resistance (mer) operon. FEMS Microbiol. Rev. 19: 239-62.
- 233. Osteen A. B. and Bibler J. P. (1991). Treatment of radioactive laboratory waste for mercury removal. Water Air Soil. Pollut. 56: 63-74.
- 234. Pahan et al (1990). "Ecological and Biochemical studies on Mercury resistant bacteria". Ind. J. Env. Hlth. 32: 250-61.

- 235. Pain A. and Cooney J. J. (1998). Characterization of organotin resistant bacteria from Boston Harbor sediment. Arch. Environ. Contam. Toxicol. 35: 412-16.
- 236. Pant A. and Singh S. P. (1995). Methyl mercury uptake by cyanobacterial immobilisates under environmental stress. PHYKOS 34: 17-25.
- 237. Pazirandeh M., Chrisey L. A., Mauro J. M., Campbell J. R. and Gaber B. P. (1995). Expression of the *Neurospora crassa* metallothionein gene in *Escherichia coli* and its effect on heavy-metal uptake. Appl. Microbiol. Biotechnol. 43: 1112-17.
- 238. Pettibone G. W. and Cooney J. J. (1988). Toxicity of methyltin to microbial populations in estuarine sediments. J. Ind. Microbiol. 2: 373-78.
- 239. Pilgrim W., Schroeder W., Porcella D. B. Santos-Burgoa C., Montgomery S., Hamilton A. and Trip L. (2000). Developing concensus: mercury science and policy in the NAFTA countries (Canada, the United States and Mexico). Sci. Tot. Environ. 261: 185-93.
- 240. Potula V. L and Hu H. (1996). Occupational and lifestyle determinants of blood lead levels among men in Madras, India. Int. J. Occup. Environ. Hlth. 2: 1-4.
- 241. Prasannakumar S., Muraleedharan P. M., Prasad T. J., Gauns M., Ramaiah N., de Souza S. N., Sardesai S. and Madhupratap M. (2002). Why is the Bay of Bengal less productive during summermonsoon compared to the Arabian Sea? Geophys. Res. Lett. 29(24): 2235, doi: 10.1029/2002GL016013.
- 242. Radstrom P., Skold G., Swedberg G., Flensburg J., Roy P. H. and Sundstrom L. (1994). Transposon Tn5090 of the plasmid R751, which carries an integron, is related to Tn7, Mu, and the retroelements. J. Bacteriol. 176: 3257-68.
- 243. Rai L. C., Malick N., Singh J. B. and Kumar H. D. (1991) Physiological and biochemical characteristics of a copper tolerant and a wild type strain in *Anabaenaa doliolum* under copper stress. J. Plant Physiol. 38: 68-74.
- 244. Rainbow P. S. and Furness R. W. Heavy metals in the marine environment. CRC Press, Inc., Florida. (1990). Pp. 1-3.
- 245. Ramaiah N. Ph.D thesis "Studies on Marine Luminous Bacteria". November (1989). Pp. 230.
- 246. Ramaiah N. and Chandramohan D (1993). Ecological and laboratory studies on the role of luminous bacteria and their luminescence in coastal pollution surveillance. Mar. Poll. Bull. 26: 190-201.
- 247. Ramaiah N., Kenkre V. D. and Verlekar X. N. (2002a) Marine environmental pollution stress detection through direct viable counts of bacteria. Wat. Res. 36: 2383-93.
- 248. Ramaiah N., Ravel J., Straube W. L. and Colwell R. R. (2002b). Entry of *Vibrio harveyi* and *Vibrio fischeri* into the viable but nonculturable state. J. Appl. Microbiol. 93: 108-16.
- 249. Ramaiah N. and De J. (2003). Unusual rise in mercury-resistant bacteria in coastal environs. Microb. Ecol. 45: 444-54.
- 250. Rani D. B. R and Mahadevan A. (1994). Cloning and expression of the mercury resistance genes of marine *Pseudomonas* sp. Strain MR1 plasmid pMR1in *Escherichia coli*. Res. Microbiol. 145: 121-27.

- 251. Rasmussen L. D. and Sørensen S. J. (1998). The effect of longterm exposure of mercury in the bacterial community in marine sediment. Curr. Microbiol. 36: 291-97.
- 252. Rasmussen L. D. and Sorensen S. J. (2001). Effects of mercury contamination on the culturable heterotrophic, functional and genetic diversity of the bacterial community in soil. FEMS Microbiol. Ecol. 36: 1-9.
- 253. Ravel J., Amoroso J. M., Colwell R. R. and Hill R. T. (1998a) Mercury-resistant actinomycetes from the Chesapeake Bay. FEMS Microbiol. Lett. 162: 177-184.
- 254. Ravel J., Schrempf H. and Hill R. T. (1998b). Mercury resistance is encoded by transferrable giant linear plasmids in two Chesapeake Bay *Streptomyces* strains. Appl. Environ. Microbiol. 64: 3383-88.
- 255. Ravel J., Elizabeth M. H. W. and Hill R.T. (2000a). Interspecific transfer of Streptomyces giant linear plasmids in sterile amended soil microcosms. Appl. Environ. Microbiol. 66: 529-534.
- 256. Ravel J., DiRuggiero J., Robb F. T. and Hill R. T. (2000b). Cloning and sequence analysis of the mercury resistance operon of *Streptomyces* sp. strain CHR28 reveals a novel putative second regulatory element. J. Bacteriol. 182: 2345-49.
- 257. Rawlins B. G., Ferguson A. J., Chilton P. J., Arthurton R. S., Rees J. G. and Baldock J. W. (1998). Review of agricultural pollution in the Caribbean with particular emphasis on small island developing states. Mar. Poll. Bull. 36: 658-68.
- 258. Reyes N. S., Frischer M. E. and Sobecky P. A. (1999). Characterization of mercury resistance mechanisms in marine sediment microbial communities. FEMS Microbiol. Ecol. 30: 273-84.
- 259. Reysenbach A. L., Giver L. J., Wickham G. S. and Pace N. R. (1992). Differential amplification of rDNA genes by polymerase chain reaction. Appl. Environ. Microbiol. 58: 3417-18.
- 260. Rippka R., Waterbury J. B. and Stainer R. Y. Isolation and purification of cyanobacteria, some general principles: *In* Prokaryotes M. P. Starr, H. Stolp, H. G. Truper, A. Balow and H. G. Schleger (eds.). Springer, New York. (1981). Pp. 212-20.
- 261. Ritter J. A. and Bibler J. P. (1992). Removal of mercury from wastewater: large scale performance of an ion exchange process. Water Sci. Technol. 25: 165-72.
- 262. Roane. T. M. and Kellogg S. T. (1996). Characterization of bacterial communities in heavy metal contaminated soils. Can. J. Microbiol. 42: 593-603.
- 263. Roane T. M., Pepper I. L. and Miller R. M. Microbial remediation of metals. In Crawford R. L. and Crawford D. L. (eds.) Bioremediation: Principles and applications. Cambridge University Press, Cambridge, UK. (1996). Pp. 312-40.
- 264. Roane T. M. and Pepper I. L. Microbial remediation of soils co-contaminated with 2, 4-dicholorphenoxy acid and cadmium. Proceedings of the 12th annual conference on hazardous research, Kansas City. (1997). Pp. 343-56.
- 265. Roane T. M. (1999). Lead resistance in two bacterial isolates from heavy metalcontaminated soils. Microb. Ecol. 37: 218-24.
- 266. Roane T. M. and Pepper I. L. (2000) Microbial Responses to Environmentally Toxic Cadmium. Microb. Ecol. 38: 358-64.

- 267. Roane T. M., Josephson K. L. and Pepper I. L. (2001). Dual-bioaugmentation strategy to enhance remediation of contaminated soil. Appl. Environ. Microbiol. 67. 3208-15.
- 268. Robinson J. B. and Tuovinen O. H. (1984). Mechanisms of microbial resistance and detoxification of mercury and organomercury compounds: physiological, biochemical and genetic analysis. Microbiol. Rev. 48: 95-124.
- 269. Rogers S. L. and McClure N. Bioremediation process optimization. In Bioremediation a critical review by Head I. M., Singleton I. and Milner G. (eds.). Horizon Scientific Press, England. (2003). Pp. 301.
- Rugh C. L., Senecoff J. F., Meagher R. B. and Merkle S. A. (1998). Development of transgenic yellow poplar for mercury phytoremediation. Nature Biotechnol. 16: 925-28.
- 271. Safe S. (1984). Polychlorinated biphenyls (PCBs) and polybrominated biphenyls (PBBs): Biochemistry, toxicology, mechanism of action. CRC Crit. Rev. Toxicol. 13: 310-95.
- 272. Sanger F., Nicklen S., Coulson A .R. (1977). DNA sequencing with chainterminating inhibitors. Proc. Nat. Acad. Sci. USA. 74: 5463-67.
- 273. Sant'ana Y. X., Chartone-Souza E. and Ferreira M. D. (1989). Drug resistance and colicinogeny of *Salmonella typhimurium* strains isolated from sewage-contaminated surface water and humans in Belo Horizonte, Brazil. Rev. Microbiol. 20 : 41-49.
- 274. Saouter E., Turner R. and Barkay T. (1994). Microbial reduction of ionic mercury for the removal of mercury from contaminated environments. Ann. N. Y. Acad. Sci. 721: 423-27.
- 275. Saouter E., Gillman M. and Barkay T. (1995). An evolution of mer-specified reduction of ionic mercury as a remedial tool of a mercury-contaminated freshwater pond. J. Ind. Microbiol. 14: 343-48.
- Sarin R., in Proceedings of the Third Indo-Dutch Workshop on Aquatic Toxicology, Centre for Environmental Studies, Anna University, Chennai, 1991, pp. 1– 29.
- 277. Sarkar A., De J. and Ramaiah N. (2003). Microbial process for degradation of PCBS in Clophen A-50 using a novel marine microorganism, *Pseudomonas* CH07. US patent no. 6,544,773 granted on April 8, 2003.
- 278. Schwarzenbach R. P., Gschwend D. M. and Imboden D. M. Environmental Organic Chemistry. New York, John Wiley & Sons, Inc. (1993). Pp. 681.
- 279. Selifonova O. V. and Barkay T. (1994). Role of Na⁺ in transport of Hg²⁺ and induction of the Tn21 mer operon. Appl. Environ. Microbiol. 60: 3502-07.
- 280. Sharma P. K., Blackwill D. L., Frenkel A. and Vairavamurthy M. A. (2000). A new *Klebsiella planticola* strain (Cd-1) grows anaerobically at high cadmium concentrations and precipitates cadmium sulfide. Appl. Environ. Microbiol. 66:3083-87.

- 281. Sherman D. F., Stroo H. F. and Bratina J. Degradation of PAH in soils utilizing enhanced bioremediation. In Gas, oil, and coal Biotechnology. Vol. 1. Akin C and Smith J. (eds.). Institute of Gas Technology, Chicago III. (1990). Pp. 417-28.
- 282. Silver S. (1996). Bacterial resistances to toxic metals- a review. Gene. 179: 9-19.
- 283. Silver S. and Misra T. K. (1988). Plasmid-mediated heavy metal resistance. Ann. Rev. Microbiol. 42: 717-43.
- 284. Silver S., Endo G. and Nakamura K. (1994). Mercury in the environment and laboratory. J. Japan. Soc. Water Envrion. 17: 235-44.
- 285. Silver S. and Phung Le T. (1996). Bacterial heavy metal resistance: New Surprise. Annu. Rev. Microbiol. 50: 753-89.
- 286. Slaveykova V. I. And Wilkinson K. J. (2002). Physiochemical aspects of lead bioaccumulation by *Chlorella vulgaris*. Environ. Sci. Technol. 36: 969-75.
- 287. Slemr F., Brunke E. G., Ebinghaus R., Temme C., Munthe J., Wängberg I., Schroeder W. and Steffen A. (2003). Worldwide trend of atmospheric mercury since 1977. Geophys. Res. Lett. 30: 1516-18.
- 288. Smith P. J. (1978). Toxicological data on organotin compounds. I.T.R.I. Publications. 538.
- 289. Smith P. J. (1980). Structure activity relationships for di- and tri-organotin compounds. I. T. R. I. Publications, 569.
- 290. Smith P. J. (1998). Chemistry of tin -II edition, Blackie academic and professionals, London, UK.
- 291. Soldano B. A., Bien P. and Kwan P. (1975). Air-borne organomercury and elemental mercury emissions with emphasis on central sewage facilities. Atmos. Environ. 9: 941-44.
- 292. Stewart C. J. and More S. J. (1990). A review of the degradation of tributyltin in the marine environment. Environ. Technol. 11: 565-70.
- 293. Strasdeit H. (1998). Methylquecksilber: eine Warnung. Nachrichten aus Chemie, Technik und Laboratorium. 46(9): 846-48.
- 294. Sturman, P. J., Steward P. S., Cunningham A. B., Bower E. J. and Wolfram J. H. (1995). Engineering scale-up of in situ bioremediation processes. A review. J. Contam. Hydrol. 19: 171-203.
- 295. Summers A. O. and Silver S. (1972). Mercury resistance in a plasmid-bearing strain of *Escherichia coli*. J.Bacteriol. 112: 1228-36.
- 296. Summers A. O. and Lewis E. (1973). Volatilization of mercuric chloride by mercury-resistant plasmid-bearing strains of *Escherichia coli*, *Staphylococcus aureus* and *Pseudomonas aeruginosa*. J. Bacteriol. 113: 1070-72.
- 297. Summers A. O. and Silver S. (1978). Microbiological transformation of metals. Ann. Rev. Microbiol. 32: 637-72.
- 298. Summers A. O. (1986). Organization, expression and evolution of genes for mercury resistance. Ann. Rev. Microbiol. 40: 607-34.

- 299. Summers A. O. (1992). The hard stuff: metals in bioremediation. Curr. Opin. Biotechnol. 3: 271-76.
- 300. Suzuki S., Fukagawa T. and Takama K. (1992). Occurrence of tributyltin tolerant bacteria in tributyltinor cadmium containing seawater. Appl. Eviron. Microbiol. 58: 3410-12.
- 301. Suzuki S., Tsukamoto K.K. and Fukagawa T. (1994). The 16S rRNA sequence and genome sizing of tributyltin resistant marine bacterium, strain M-1. Microbiology. 101: 101-09.
- 302. Taylor and Francis. Lead pollution (pp111-114), Mercury and methylmercury (pp118-121), Cadmium (pp76-77). In: Timbrell J. A. Introduction to toxicology. 2nd edition. London (1995).
- 303. Thomas D. L.; Montes J. G (1978). Spectrophotometrically assayed inhibitory effects of mercuric compounds on *Anabaena flosaquae* and *Anacystis nidulans* (Cyanophyceae). J. Phycol. 14: 494-99.
- 304. Tisdale W. H. (1943). British patent no. 578.312, UK.
- 305. Trevors J. T. (1986). Mercury resistant bacteria isolated from sediment. Bull. Environ. Contam. Toxicol. 36: 405-11.
- 306. Unterman, R. A. History of PCB biodegradation. Bioremediation: Principles and Applications. R. L. Crawford and D. L. Crawford. Great Britain, Cambridge University Press. (1996). Pp. 209-53.
- 307. Viney I. and Bewly R. J. F. (1990). Preliminary studies on the development of microbiological treatment for polychlorinated biphenyls. Arch. Environ. Contam. Toxicol. 19: 241-52.
- 308. Volesky B. and Holan Z. R. (1995). Biosorption of heavy metals. Biotechnol. Prog. 11: 235-50.
- Wagner-Döbler I., Lünsdorf H., Lübbehüsen T., Von Canstein H. F., and Li Y. (2000). Structure and Species Composition of Mercury-Reducing Biofilms. Appl. Environ. Microbiol. 66: 4559-63.
- 310. Wagner-Döbler I. (2003). Pilot plant for bioremediation of mercury-containing industrial wastewater. Appl. Microbiol. Biotechnol. 62: 124-33.
- Wang C. L., Michels P. C., Dawson S. C., Kitisakkul S., Bross J. A., Keasling J. D. and Clark D. S. (1997). Cadmium removal by a new strain of *Pseudomonas* aeruginosa in aerobic culture. Appl. Environ. Microbiol. 63: 4075-78.
- 312. Wang Y., Mahler I., Levinson H. S., Silver S. and Halvorson H. O. (1987). Cloning and expression in *Escherichia coli* of chromosomal mercury resistance genes from a *Bacillus* sp. J. Bacteriol. 169: 4848-51.
- 313. Wang Y., Moore M. J., Levinson H. S., Silver S. and Walsh C. T. (1989). Nucleotide sequence of a chromosomal mercury resistance determinant from a *Bacillus* sp. With broad-spectrum mercury resistance. J. Bacteriol. 171: 83-92.
- 314. Weisberg W. G., Barns S. M., Pelletier D. A. and Lane D. J. (1991). 16S ribosomal DNA amplification for phylogenetic study. J. Bacteriol. 173: 697-703.

- 315. White J. S., Tobin J. M. and Cooney J. J. (1999). Organotin compounds and their interaction with microorganisms. Can. J. Microbiol. 45: 541-54.
- 316. Wiegel J. and Wu Q. (2000). Microbial reductive dehalogenation of polychlorinated biphenyl. FEMS Mirobiol. Ecol. 32: 1-15.
- 317. Williams F, Robertson R. and Roworth M. Scottish. Centre for Infection and Environmental Health: Detailed profile of 25 major organic and inorganic substances. 1st ed. Glasgow: SCEIH. (1999).
- 318. Williams J. W. and Silver S. (1984). Bacterial resistance and detoxification of heavy metals. Enzyme Microb. Biotechnol. 6: 530-37.
- 319. Wilson J. R., Leang C., Morby A. P., Hobman J. L. and Brown N. L. (2000). MerF is a mercury transport protein: different structures but a common mechanism for mercuric ion transporters? FEBS Lett. 472: 78-82.
- 320. Winfrey M. R. (1985). The role of methane producing and sulfate reducing bacteria in mercury methylation. Abstr. Ann. Meet Amer. Soc. Microbiol. Q79: 271.
- 321. Winfrey M. R. and Rudd J. W. M. (1990). Environmental factors affecting the formation of methylmercury in low pH lakes. Environ. Toxicol. Chem. 9: 853-69.
- 322. Wood J. M., Kennedy F. S.and Rogen C. G. (1968). Synthesis of methyl mercury compounds by extracts of a methylogenic bacterium. Nature (London). 220: 173-74.
- 323. Wuertz S., Miller C. E., Pfister R. M. and Cooney J. J. (1991). Tributyltinresistant bacteria from estuarine and freshwater sediments. Appl. Environ. Microbiol. 57: 2783-89.
- 324. Xiao Z., Sommar J. and Lindquist O. (1998). Atmospheric mercury deposition on Fangjing mountain nature service, Guizhou, China. Chemosphere. 36: 2191-200.
- 325. Yadav J. S., Reddy C. A., Quensen J. F. and Tiedje J. M. Degradation of polychlorinated biphenyl mixtures in soil using Phanerochaete chrysosporium in nutrient rich, non-ligninolytic conditions. USP no. 6,107,079. August 22, 2000.
- 326. Yamada H., Takayanagi K., Tateishi M., Tagata H. and Ikeda K. (1997). Organotin compounds and chlorinated biphenyls of livers in squid collected from coastal waters and open oceans. Environ. Poll. 96: 217-26.
- 327. Zeroual Y., Moutaouakkil A. and Blaghen M. (2001). Volatilization of mercury by immobilized bacteria (*Klebsiella pneumoniae*) in different support by using fluidized bed reactor. Curr. Microbiol. 43: 322-27.

Web links:

- 1. http://arcweb.sos.state.or.us/rules/0103_Bulletin/0103_ch603_bulletin.html
- 2. http://in.rediff.com/money/2003/nov/04mercury.htm
- 3. http://www.ebi.ac.uk
- 4. http://www.epa.gov
- 5. http://www.inchem.org/documents/ehc/ehc/ehc086.htm#SectionNumber:3.3.

- 6. http://www.inchem.org/documents/ehc/ehc/ehc140.htm
- 7. http://www.mmjp.or.jp/JOF/chikyukan/2003/5/5_2_7/e_1.html
- 8. http://www.thaiunique.com/varian/doc/ICP-Lib2.pdf